



Bay of Plenty Coastal Water Quality, 2003 - 2004

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Cover Photo: Looking out to Karew a and Tuhuna (Mayor) Islands from Matakana Island beach. Photo by Sam Stephens.

Executive Summary

A study of the central Bay of Plenty's coastal shelf waters was undertaken to collect data for the support of modelling and estimating the sustainability and carrying capacity of aquaculture. The study included physical and chemical analysis of the waters, quantification of phytoplankton communities, current measurements and temperature profiling of the water column and the use of remote sensing to provide a synopsis of seasonal and spatial patterns of sea surface temperature and chlorophyll-a concentrations. In this report the data gained from physical, chemical and phytoplankton analysis of the water column is presented.

Field work to collect physical, chemical and phytoplankton data was conducted along three transects spanning the coastal shelf out to a depth of 200m at Pukehina, Whakatane and Opotki. Sampling was also conducted from October 2003 through to May 2004. An additional instrument (CTD) survey was conducted in December 2004 across a wider area of the Bay of Plenty, out tow ards the east.

Results are similar to the earlier findings of the coastal shelf survey undertaken in 1996/97 in respect of the major seasonal and depth related trends observed in the water column. In this survey the use of a wider range of monitoring equipment to measure many of the parameters means that there is a far better resolution and understanding of the oceanographic processes operating in these waters. The CTD and current measurements show that the shelf waters area complex system with wide variations around the Bay of Plenty at any one point in time.

Contents

Acknowledgements1			
Exec	iutive Summaryi		
Chap	oter 1: Introduction1		
1.1	Scope1		
1.2	AMA Project background2		
Chap	oter 2: Methods		
2.1	Location and Physical Environment5		
2.2	Sampling Programme6		
2.3	Methods7		
Chap	oter 3: Results		
3.1	Physical Attributes9		
3.2	Nutrients25		
3.3	Biological		
Chapter 4: Summary			
Chap	oter 5: References		
Арре	andices		
Apper	ndix 1 – Site Location and Sampling Program		
Apper	ndix 2 – BOP Offshore Transect Survey Results41		
Apper	ndix 3 – CTD Profiles From Transect Surveys47		

List of Tables

Table 1	Methods used for chemical / biological analysis7
Table 2	Gridding methods used in the plotting of data8
Table 3	Phytoplankton species list and the seasonal abundance rank obtained from the three transects sampled in Bay of Plenty coastal waters
Table 4	Overall percentage of diatoms cells in the total number of phytoplankton cells counted from spot samples on each transect
Table 5	Total numbers of phytoplankton cells counted in spot samples from the Pukehina transect
Table 6	Total numbers of phytoplankton cells counted in spot samples from the Whakatane transect
Table 7	Total numbers of phytoplankton cells counted in spot samples from the Opotiki transect

List of Figures

Figure 1	Location of transect sampling sites and the wave buoy in the Bay of Plenty coastal shelf waters
Figure 2	Secchi depth and vertical light extinction coefficients recorded along each of the transects from October 2003 to May 2004
Figure 3	Temperature (°C) plots from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf13
Figure 4	Salinity (psu) plots from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf15
Figure 5	Density (σ_t) plots from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf16
Figure 6	Dissolved Oxygen (%) plots from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf17
Figure 7	Turbidity (NTU) plots from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf
Figure 8	Suspended solids (g/m ³) plots from Opotiki, Whakatane and Pukehina transects n the BOP continental shelf20
Figure 9	Photosynthetically Active Radiation plots from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf21
Figure 10	Chlorophyll-a plots (as fluorescence measurements) from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf

Figure 11	Concentration of oxidised nitrogen from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf	. 23
Figure 12	Concentration of dissolved reactive phosphorus from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf	. 24
Figure 13	Relationship betw een Dissolved Reactive Silica and conductivity	. 25
Figure 14	Relationship betw een instrument fluorescence values and Chl-a (mg/m ³)	. 27

1.1 **Scope**

This report presents the results of field survey work jointly conducted by Environment Bay of Plenty and ASR Limited staff along a set of transects spanning the Bay of Plenty coastal shelf w aters. The work was part of the Aquaculture Management Areas (AMA) science project, collecting physical and biological data (Oct 2003 – December 2004) to support assessment of the productivity and sustainability of coastal shelf w aters within the Bay of Plenty. The specific objectives of the projects field surveys were to;

- Provide quantitative data on the spatial and seasonal abundance of phytoplankton species.
- Assess spatial and seasonal patterns of nutrients required for phytoplankton grow th.
- Assess spatial and seasonal patterns of physical parameters.
- Measurement of current direction and velocity.
- Improve our understanding of the oceanographic processes and ensure data collection that will support modelling of the shelf system.

The magnitude of the effort and the success with the field measurements is a creditto all staff involved.

Some aspects of the field surveys or other components of the overall project, such as the current and wave data will be reported separately. Other reports being produced by ASR Ltd (Longdill *et.al* 2005a & b, Beamsley *et.al* 2005, Mead *et.al* 2005) on the field data summarise:

- Measurements of currents and temperatures. 12 months of intermittent Acoustic Doppler Current Meter and thermistor records were collected during the 2-year programme.
- Observations with seabed video over the shelf with grain size, mud contents and biological organism assessments.

1.2 **AMA Project Background**

Aquaculture as an industry in New Zealand has been growing rapidly in recent years and has become a substantial producer and export earner. The industry is still largely dominated by green lipped mussel and salmon farming but has the potential to produce a very wide range of product. In some areas such as the Marlborough Sounds suitable farming space has become limited and is considered a valuable asset. In the Bay of Plenty only around 4 ha of aquaculture existed up until May 2001 at which time it then received tw o large applications for a total of 8,760 ha.

In March 2002 the government imposed a moratorium on the granting of marine space for the purposes of aquaculture due to the explosion of demand and the complicated and sometimes adhoc approach to the processing of consents. The moratorium w as "to provide Regional Councils with the opportunity to plan for Aquaculture Management Areas". It was also intended to remove much of the dual process that currently exists to make it more straight forw ard and easier for industry to establish.

The moratorium has now meant that Regional Councils must work out where to allow aquaculture to take place and how much. Environment Bay of Plenty is dealing with this task under both a science and planning project. The planning project is defining areas that will be most suitable. The approach taken is to define and map the various values of the marine environment that may potentially clash with aquaculture. This includes ecological, cultural, recreational fishing, commercial fishing, recreational activities, navigation, landscape and water quality values and issues.

Aquaculture can put considerable pressure on some components of the marine environment. Mussel farming for instance can cause phytoplankton depletion, change plankton species composition, result in nutrient changes, deposition of shell and faeces, impacts on water quality and a range of other actual or potential impacts. One of the most important questions is that of sustainability from a productivity view point. For example how much mussel farming could the Bay of Plenty coastal shelf waters support without such farms depleting phytoplankton levels to an extent that other inshore shellfish beds may be impacted, inhibiting their ability to grow at normal rates and sustainable levels.

The aquaculture industry grow th and moratorium has imposed an urgent need to obtain a sound understanding of the oceanographic processes, productivity and carrying capacity of the Bay of Plenty coastal w aters. The science project being run as part of the w hole AMA project is intended to address many of the issues and know ledge gaps mentioned above. It w as started in July 2003 and w ill be concluded at the end of 2005 w ith a final report in March 2006.

The science project in its first year (2003/4) aimed to set up a wave buoy, current meter and gather data from field surveys on the physical and biological status of the coastal waters during each season of the year. In addition remote sensing data is being used to assist in defining the seasonal and spatial status of Bay of Plenty waters for sea surface temperature and chlorophyll-a levels. Further work on defining the nature of the seabed and benthic communities will also be undertaken.

A large component of the science project will be the development of hydrological and productivity models of the Bay of Plenty. Plankton, nutrient, physical and remote sensing

data will all help in developing and calibrating numerical models aimed at estimating impacts, carrying capacity and sustainability of marine farming. Overall the project is primarily aimed at providing information needed to assist Environment Bay of Plenty's planning responsibilities in a way which will also help to safeguard the regions natural marine resources and ecosystems.

2.1 Location and Physical Environment

Field sampling for the AMA science project builds upon a previous survey of the coastal shelf w aters undertaken in 1996/97 (Park 1998). In Figure 1 below the three transects in the centre of the bay (Pukehina, Whakatane & Opotiki) running from shore (10m depth) out to the edge of the continental shelf (200m depth) w ere used for the 2003/2004 field surveys. The Tauranga and Whakatane transects w ere both used in the 1996/97 survey. Grid references for the sampling points along each of these transects are provided in Appendix 1.



Figure 1 Location of transect sampling sites and the wave buoy in the Bay of Plenty coastal shelf waters

The Bay of Plenty coastline is exposed to wind and waves from the northwest to easterly directions and is a lee shore in respect of New Zealand's prevailing southwesterly wind flows. Offshore waters are predominantly influenced by the East Auckland Current, which flows south-eastward along the north-east coast of the North

Island to East Cape where the main core of the current turns north-eastward (Ridgway and Greig 1986). Influence of the current is thought to be variable with limited penetration inshore.

Inshore currents are highly variable responding to both tidal effects and wind forcing (Healy et al 1988, Harms 1989). During periods of high wind strengths, currents were observed to flow in the direction of the wind with velocities of up to 40 cm sec⁻¹. In addition, periods of onshore bottom current occurred with offshore wind and the reverse with onshore winds. Residual current flows during periods of weak and variable winds appear to be to the south, possibly as an inshore influence of the East Auckland Current. Similar residual currents have been observed from a long-term current observation on the east Coromandel shelf (Bradshaw et al 1991). Initial results collected from this project would also support this synopsis.

The inshore end of the Tauranga transect is influenced by tidal currents and flushing associated with the southern entrance of Tauranga Harbour. A similar situation exists with the Whakatane transect and estuary.

The southern entrance to Tauranga Harbour has a recorded flow of 153×10^6 m³ with maximum current speeds of 2.65 m.s⁻¹ (Beca Carter Hollings and Ferner Ltd 1978). Freshw ater inflow s to the southern harbour have been estimated to be around 30.5 ms⁻¹ (<1% of tidal compartment) with the Wairoa River contributing 58% of the total (McIntosh 1994). The catchment of the northern Tauranga Harbour basin is much smaller with total freshw ater inflow of around 4.6 m³s⁻¹.

In contrast to the Tauranga situation Whakatane Estuary is much smaller with a high riverine influence. The Whakatane River, which flow s into the estuary, has a mean flow of 57 m²s⁻¹. Peak ebb-tide flow s from the estuary have been gauged at 140 m²s⁻¹ during river flow s of 18 m²s⁻¹ (Healy 1983). Catchment soils are still influenced by volcanic ash but most soils are derived from w eathering of the underlying greyw acke rocks.

Offshore of Opotiki the situation is similar to that of Whakatane, especially regarding the catchment geology. Opotiki Estuary is even smaller measuring only 0.6 km². The freshwater inflow from the Waioeka and Otara Rivers combined is also slightly less with a mean annual flow of 44m³ s⁻¹. Both these rivers flow through Opotiki Estuary.

At the Pukehina transect there is not a lot of freshwater influence even at the inshore 10m depth site. The Waitahanui Stream is around 1 km away and has a mean annual flow of 6 m³ s⁻¹. The Kaituna River is 15 km to the northwest and has a mean annual flow of 39 m³ s⁻¹, but due to the distance it has only a very diffuse effect on salinity.

2.2 Sampling Programme

Field sampling was conducted at sites along each of the Pukehina, Whakatane and Opotiki transects show n in Figure 1. Surveys were conducted four times within a year to obtain data on seasonal variations in water quality and plankton assemblages. Each transect has sampling sites set at the 10, 20, 30, 50, 100 and 200m depth contours. This takes each transect from within a kilometre of the shore out to the edge of the continental shelf. This ensured that neritic coastal waters were fully covered and allow s comparison to oceanic water masses.

Samples were obtained on all transects within a day of the following dates:

17th October, 2003, 3rd December 2003, 18th March 2004, 25th May 2004, 1st August 2004.

An additional set of transect were surveyed with CTD instruments on 7-9th December 2004.

At each of the sampling sites instrument readings, water samples, bacterial samples, and plankton samples were taken at specified depths. Appendix 1 sets out the sampling programme in detail listing depths and all chemical, physical, and biological sampling conducted.

The surface sample (0-5m) for all chemistry and plankton samples taken from each sampling location was obtained as a depth integrated tube sample. All other samples from each of the specified depths were point samples taken with a 3 litre van Dorn bottle.

Instruments readings at each site w ere taken w ith a SBE 19plus SEACAT Profiler from the surface down to the seabed. This probe recorded temperature, conductivity, pressure (depth), Oxygen (SBE 43), PAR (LI-COR LI-193SA), fluorometer (Turner SCUFA), and OBS (turbidity – Turner SCUFA). In addition temperature and conductivity w ere recorded from the w ater samples retrieved w ith the van Dorn bottles for sample analysis using a hand held YSI meter.

2.3 Methods

The following methods were used to derive the results from the field sampling. All samples for chemical analysis were stored and returned with the time period stipulated according to the method requirements;

Parameter	Method	Detection Limit [†]
Suspended Solids	APHA method 2540D	0.1 g/m ³
Total Organic Carbon	catalytic oxidation, IR detection. APHA 5310B 20 th ed.	0.5 g/m ³
Dissolved Organic Carbon	0.45 µm nylon filter, catalytic oxidation, IR detection	0.5 g/m ³
Dissolved Reactive Silica	On-site filtration of sample. Moly bdosilicate/ascorbic	1 mg/m ³
Dissolved iron	On-site filtration of sample. 0.45 µm filtered sample.	4 mg/m ³
Total nitrogen	persulphate digestion, auto cadmium reduction, flow	1 mg/m ³
Ammonium nitrogen	NWASCO Misc Pub. No. 38, 1982. phenolhy pochlorite	1 mg/m ³
Oxidised nitrogen	flow injection analyser, APHA 4500 NO3-1	1 mg/m ³
Total Phosphorus	acid persulphate digestion, molybdate colorimetry. Flow	4 mg/m ³
Dissolved Reactive Phosphorus	NWASCO Misc Pub. No. 38, 1982. Antimony –	4 mg/m ³
Dissolved non-		0.5 g/m ³

Table 1 - Methods used for chemical / biological analysis.

Non-purgable organic	acid pre-treatment to remove inorganic carbon.	0.5 g/m ³			
Secchi disc	measured in metres (to 0.1m increments) without a				
Vlec	vertical light extinction coefficient derived from PAR measurements (LI-COR LI-193SA) for the surface photic				
Enterococci	APHA method 9230c - number cf u/100ml				
рН	pH measurement @ 25°C				
Chlorophyll-a total filterable	total phytoplankton filterable on 0.7µm and above, GFC filtration, acetone pigment extraction,	0.1 mg/m ³			
Nano-chl-a	as for total Chlorophyll-a but only for plankton 0.7-20µm	0.1 mg/m ³			
Micro-chl-a	as for total Chlorophyll-a but only for plankton 20-200	0.1 mg/m ³			
Phytoplankton	Phytoplankton samples were collected from set depths using a van Dorn bottle with the exception of the surface (0-5m) sample which was obtained from a depth integrated tube sample. Around 250 ml of sample was preserved with Lugol's Iodine. These samples were then cont to NUWA in Wollington for applying by other Hop				
[†] Detection limit with 95% confidence, some results are below this level					

2.3.1 Contour Plot Methods

The raw data from both the CTD (data at each meter down the water column) and also from the water samples (data more sparse at discrete intervals – see Appendix 1) were plotted using Golden Software's Surfer software. Details of the gridding methods and dimensions are tabulated in Table 2.

Table 2	Gridding methods used in the plotting of data.
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	Transect	Grid extent	Gridding method	Grid cell size
CTD Data	Opotiki	0-38000 m(x), 0210 m(y)		
	Whakatane	0-31000 m(x), 0210 m(y)	Krigging	300 m(x) x 3 m(y)
	Pukehina	0-25000 m(x), 0210 m(y)		
	Opotiki	0-38000 m(x), 0210 m(y)	Triangulation with	
Chemical Data	Whakatane	0-31000 m(x), 0210 m(y)	linear	3000 m(x) x 15 m(y)
	Pukehina	0 – 25000 m(x), 0210 m(y)	interpolation	

3.1 Physical Attributes

3.1.1 Water Clarity

Results for measurements of Secchi depth and vertical light extinction co-efficient (Vlec) are displayed in Figure 2. All the transects, which ran out from Pukehina, Whakatane and Opotiki, show ed a number of consistent trends as expected. All have a significant improvement in water clarity out to the edge of the continental shelf. Inshore (10 mdepth contour) Secchi depth measurements ranged from 1 - 7 m while offshore values show ed a larger degree of variation. At the 200 m contour visibility reached a maximum of about 30 m in the autumn (March) sampling period.

Overlaid on the inshore offshore trend is the seasonal component. This was most easily seen further offshore at the 200 m contour where there were measurements of only 6-7 m in spring and early summer compared to the autumn maximum of 30 m.

The overall timing and magnitude of the inshore–offshore and seasonal variations are similar to the previous survey that was conducted in 1996/97(Park 1998).

3.1.2 Salinity/conductivity

Salinity profiles for the Pukehina, Whakatane and Opotiki transects are presented in Figures 4 and 5. In addition the profile of the water column at each sampling pointalong the transects is given in Appendix III.

The predominant feature seen in all plots is the seasonal influence of freshw ater input into the Bay of Plenty from land. In the profile plots in Appendix III, there are many examples where salinity changes markedly with depth in association with the different layers of w ater. Many of these distinct w ater layers show reversal of current direction (Beamsley et.al 2005). The seasonal pattern is very similar to the findings of a similar survey undertaken in 1996/97 (Park 1998).





3.1.3 Temperature

Temperature profiles for the Pukehina, Whakatane and Opotiki transects are presented in Figures 3 providing a visual perspective across the shelf waters of each transect. All transects show the strong development of a seasonal thermo cline in which stratified surface waters are much warmer than the deep water. In summer water temperatures are typically 20 –22 $^{\circ}$ C at the surface with a thermo cline extending from 20 – 50 m depth with at least 2 degrees of temperature variation with the underlying water. The thermo cline tends to increase in depth further out from shore.

Winter water temperatures drop to around 12 - 13 °C. In the May 2004 survey of the transects, the surface waters were cooling with the deeper water being slightly warmer. At the edge of the shelf (200m contour) the deep oceanic water could usually be detected and measured around 12.5 at this depth regardless of season.

As shown by the CTD profile plots in Appendix III, temperature can change quite sharply in association with more distinct layers of water. As mentioned above for salinity, some of these water layers are also moving in different directions.

3.1.4 Suspended Solids and turbidity

Figures 7 and 8 present turbidity and suspended solids data. Density of sample points for suspended solids is not high so some of the results appear a little bit variable. Some of the trends noted in previous surveys (Park 1998) are apparent. That is for higher levels of suspended solids to occur both inshore and seasonally in w inter and spring.

3.1.5 Dissolved oxygen

Dissolved oxygen profiles (% saturation) for the Pukehina, Whakatane and Opotiki transects are presented in Figure 6 providing a visual perspective across the shelf waters of each transect. A more detailed depth profile relationship between dissolved oxygen and the physical properties of the shelf waters is provided by the graphics in Appendix III.

The general pattern show n by dissolved oxygen is for a seasonal peak to occur in the surface waters in spring and early summer. High concentrations of dissolved oxygen clearly relate to the amount of phytoplankton present. There is also some association of dissolved oxygen levels and the physically separate water bodies which are often present. Figure 6 also shows clearly a high demand for oxygen associated with the seafloor, particularly in the mid-shelf region.



Figure 3 Temperature (°C) plots from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf.



Figure 4 Salinity (psu) plots from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf.



Figure 5 Density (σ_t) plots from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf.



Figure 6 Dissolved Oxygen (%) plots from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf.



Figure 7 Turbidity (NTU) plots from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf.



Figure 8 Suspended solids (g/m³) plots from Opotiki, Whakatane and Pukehina transects n the BOP continental shelf.



Figure 9 Photosynthetically Active Radiation plots from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf.

21



Figure 10 Chlorophyll-a plots (as fluorescence measurements) from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf.



Figure 11 Concentration of oxidised nitrogen from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf.

23



Figure 12 Concentration of dissolved reactive phosphorus from Opotiki, Whakatane and Pukehina transects on the BOP continental shelf.

3.2 Nutrients

Results for all the nutrient analyses undertaken along the transects is provided in Appendix II. Some of the data has been graphically presented to highlight observed patterns.

3.2.1 Dissolved Reactive Silica

The previous study of the Bay of Plenty coastal waters in 1996/97 established a detectable influence of freshwater inputs on Dissolved Reactive Silica levels along the open coast. Closer to shore levels tend to be higher and there is also a seasonal pattern with levels being low er in summer and autumn when freshwater inflows are low er. The general relationship between conductivity and Dissolved Reactive Silica is shown in Figure 13 below.





3.2.2 **Dissolved Reactive Phosphorus**

Contour plots of dissolved reactive phosphorus concentrations out across each transect for the seasonal sampling runs is provided in Figure 12. These plots show that all transects displayed very low concentrations in late summer/early autumn. There were consistently high concentrations in the deep water near the edge of the coastal shelf.

Concentrations of dissolved reactive phosphorus were also consistently higher in the bottom waters across the shelf, which points to replenishment from either the sediments or from deep oceanic water up-welling in across the coastal shelf.

3.2.3 Total Oxidised nitrogen

Contour plots of total oxidised nitrogen concentrations out across each transectfor the seasonal sampling runs are provided in Figure 11. Data is missing for the December 2003 runs as all the samples were lost in the laboratory.

Overall trends are similar to the earlier Bay of Plenty study (Park 1998) with a seasonal pattern of depletion in summer within the upper stratified waters. There is also consistently higher concentrations in the bottom waters, particularly near the edge of the coastal shelf. As with dissolved reactive phosphorus, this points to sediment regeneration and up-w elling of the deep oceanic water in across the shelf.

3.3 Biological

3.3.1 Chlorophyll-a

Concentrations of chlorophyll-a measured during the year on each of the transects is best show n graphically by results presented in Figure 10 and the water columprofiles graphed and presented in Appendix III.

During the course of the field surveys water samples were collected from set points and analysed for chl-a concentrations to assist with both calibration and general validation of instrument readings (see Appendix II). Calibration of the fluorometer is based on 29 samples collected on the 22nd October and 9th December 2005. *In vivo* estimates of chlorophyll concentration were corrected for turbidity effects using a multiple regression model as follow s:

 $Y = m_x x + m_z z + b$, result obtained for this regression was an r² of 0.85877

Where:

Y = corrected chlorophyll value (mg/m^3)

 m_x = coefficient (slope) for *in vivo* chl-a = 0.82568

 m_z = coefficient (slope) for turbidity = -0.2295

b = y intercept = -0.41137

Throughout most of the transect surveys turbidity was generally very low so a simpler equation of y (chl-a mg/m³) = 0.82568 * fluorescence value - 0.41137 could be used to convert the fluorescence data. The simpler relationship betw een just fluorescence and Chlorophyll-a (mg/m³) is shown in Figure 14 below.



Figure 14 Relationship between instrument fluorescence values and Chl-a (*mg/m*³).

The highest fluorescence values from the survey were obtained on the 17th October 2003 at the 50 m depth contour on the Whakatane transect. The peak fluorescence value of 40 occurred as a pronounced spike in a very narrow depth range at about 17 m. This maximum converts to just over 30 mg/m³ of chl-a w hich is very high. Generally peak spring values as measured by fluorometer appeared to be around 18 mg/m³ of chl-a, w hich is still indicative of high productivity.

Chlorophyll-a concentrations at all sampling sites tended to peak in spring. During summer and autumn measured levels were much low er at around 2 mg/m^3 .

In the mid shelf area (50 - 100 m depth) the depth at which the chl-a peaks occurred, show ed a seasonal change associated with the development of the thermocline. The peak was deepest in summer when a strong and deep thermocline was also present.

3.3.2 **Phytoplankton species**

Results of the phytoplankton enumeration and identification show ed a number of typical trends. Table 3 below contains the list of species identified from the entire survey of the transects. In spring the most abundant species w as the diatom *Eucampia zoodiacus*, w hile the dinoflagellate *Protoperidinium* spp. dominated in summer, then in autumn it w as *Nitzschia* spp. follow ed by *Chaetoceros* spp. in w inter. The overall pattern w as that the dinoflagellates w ere more abundant in summer and autumn. This is show n in Table 4 below w hich provides the seasonal percentage composition of diatoms in the phytoplankton counts. All three transects show that numerically the diatoms w erevery dominant in spring.
Table 3

Phytoplankton species list and the seasonal abundance rank obtained from the three transects sampled in Bay of Plenty coastal waters

Diatoms	Spring	Summer	Autumn	Winter
Asterionella glacialis	19		17	6
Cerataulina spp.				15
Cerataulina pelagica	17			
Chaetoceros spp.	5	13	12	1
Coscinodiscus sp.	21		24	27
<i>Detonula</i> spp.	2	8	11	10
Diploneis sp.		15	15	28
Ditylum brightwellii	10			
Ditylumspp.				16
Eucampia zoodiacus	1			
<i>Eucampia</i> sp.		16	14	17
Fragilariopsis sp.		19		
Guinardia spp.	8			19
Lauderia annulata	16			
Lauderia sp.				9
Navicula spp.	9	6	9	14
Nitzschia longisimma	6			
<i>Nitzschia</i> spp.		2	1	8
Odontella mobiliensis	7			
Odontella sp.				7
Pleurosigma spp.		17	21	18
Pseudonitzschia spp.	4	5	4	3
Rhizosolenia imbricata	13			21
Rhizosolenia stolterforthii	12			5
Rhizosolenia spp.		12	20	13
Skeletonema costatum			8	2
Stephanopyxis sp.				11
Thalassosira spp.	3	4	7	4
Dinoflagellates				
Alexandriumsp.			6	22
Ceratium furca	22			32
Ceratium tripos	2	14		29
Ceratium spp.			19	30
Dinophysis sp.			18	31
Gymodinium spp.		10	16	25
Gyrodinium spp.	14	9	13	26
Oxytoxumsp.			22	
Polykrikos sp.		3		33
Prorocentrum spp.		11	5	20
Protoperidiniumspp.	18	1	10	23
Pyraminmonas sp.		18		
Scrippsiella sp.			2	24
Non-photosynthetic species	11			
Unknown dinof lagellates (25 micron)	15			

Micro flagellates			
Cryptomonads	7	3	12
Silicoflagellate			
Dictyocha sp.		23	

Table 4Overall percentage of diatoms cells in the total number of phytoplankton
cells counted from spot samples on each transect.

	Spring	Summer	Autumn	Winter
Pukehina	98	57	56	95
Whakatane	99	27	56	85
Opotiki	98	23	49	93

Results of the numbers of phytoplankton cells present in each of the samples are given in Tables 5-7 below. On each of the transects the seasonal trend in total cell numbers is very similar. Numbers are highest in spring and decline to the low est levels in autumn.

In terms of spatial distribution there is a pattern of higher phytoplankton cell numbers occurring inshore. How ever in the spring transect sampling the numbers arejust as high w ell offshore at the 200 m depth contour. The highest recorded cell numbers in any sample w as from 20 m depth at the 200 m depth contour around 35 km offshore from Opotiki.

		Oct 03	Dec 03	Mar 04	May 04		
Contour	depth (m)	Number of phytoplankton cells (1,000's/l)					
10	0		200	31	300		
20	0	122	135	10	186		
20	20	60	55	14	146		
30	0	210	13	25	164		
30	20		10	9	120		
50	0	234	73	9	223		
50	20	286	2	2	151		
50	30	521	17	6	109		
50	50	100	2	10	43		
100	20				5		
100	30				13		
100	100	8	7	2	14		
200	0	326	81	4			
200	20	485	86	3			
200	30	543	134	5			
200	50	121	2	3			
	mean	251	58	9	123		

Table 5	Total numbers of phytoplankton cells counted in spot samples from the
	Pukehina transect.

		Oct 03	Dec 03	Mar 04	May 04			
Contour	depth (m)	Nun	Number of phytoplankton cells (1,000's/l)					
10	0	105	46	17	208			
20	0	159	58	11	219			
20	20	32	6	23	34			
30	0	139	36	3	165			
30	20	481	31	18	273			
50	0	203	670	10	90			
50	20	409	24	5	72			
50	30	171	41	14	40			
50	50	25	3	17	5			
100	10				2			
100	20				4			
100	30				1			
100	100	15	1	1	1			
200	0	309	9	4				
200	20	250	2	8				
200	30	280	4	2				
200	50	15	1	17				
	mean	185	67	11	85			

Table 6Total numbers of phytoplankton cells counted in spot samples from the
Whakatane transect.

Table 7	Total numbers of phytoplankton cells counted in spot samples from the
	Opotiki transect.

		Oct 03	Dec 03	Mar 04	May 04
Contour	depth (m)	Num	ber of phytopl	ankton cells (1,0	00's/I)
10	0	43	20	7	109
20	0	151	37	13	224
20	20	22	7	6	104
30	0	159	69	3	113
30	20	601	6	2	201
30	30		6		
50	0	311	55	2	92
50	20	313	39	5	46
50	30	53	19	8	125
50	50	3	6	18	55
100	20				10
100	30				3
100	100	8	1	1	4
200	0	352	1	2	
200	20	672	6	2	
200	30	443		3	
200	50	189	4	4	
	mean	237	20	5	91

Chapter 4: Summary

Results from this survey program have provided a wealth of information on the Bay of Plenty coastal waters. Observed seasonal and spatial trends out across the continental shelf are very similar to those observed in 1996/97. The use of CTD profiling equipment and fluorescence probe has improved the understanding of the vertical structure of the water column. The data will be used to help calibrate a model the physical dynamics of the Bay of Plenty coastal shelf system

Current records have shown a complex system on the Bay of Plenty continental shelf being driven primarily by wind with tides and external currents (East Auckland Current) also having an influence (Beamsley et al 2005). This confirms the earlier findings of Healy et al (1988).

The major physical patterns within the Bay of Plenty coastal waters are the seasonal changes in water temperature with associated summer thermo cline development. Winter temperatures are usually around 12 - 13 °C while in summer they reach 21 - 22 °C. Nutrient levels are generally higher in winter and become depleted in the surface waters over summer. As in the earlier study of 1996/97 (Park 1998) data suggests that nutrient recycling from the sea floor and inputs from the deep oceanic water via up-w elling are both important to overall productivity. Water clarity also improves seasonally in late summer and is highest out away from the shore. Influence of terrestrial freshwater inputs and associated nutrient and clarity factors is highest in winter near the coast, often forming very large surface plumes during the larger river fresh or flood events.

Phytoplankton levels are generally highest in late winter and spring showing a pattern of spatial decline out across the shelf. Peak concentrations of around 670,000 cells per litre were recorded from water samples during the surveys but peak levels are likely to be much higher. Datoms tend to dominate during winter and spring while dinof lagellates are proportionately more abundant in summer.

Fluorometer measurements show that estimated chlorophyll-a concentrations (another measure of phytoplankton productivity) reached peaks of around $15 - 18 \text{ mg/m}^3$. The highest recorded estimate of chlorophyll-a concentration was a very sharp peak of around 40 mg/m^3 . During summer fluorometer estimates of maximum chlorophyll-a concentrations tend to be around $1-2 \text{ mg/m}^3$. Large and consistent spatial variations also occur across the Bay of Plenty depending on the prevailing w eather/current conditions. During the last survey in December 2004 chlorophyll-a concentrations to the east of Whakatane w ere low right across the shelf $(1 - 2 \text{ mg/m}^3) \text{ while the areas to the w est show ed moderate levels } (5 - 10 \text{ mg/m}^3)$.

This survey and report form part of the total AMA science project for the Bay of Plenty, which includes work on the currents and temperatures, sediments and biology. It also includes an analysis of remote sensing data to provide a synopsis of seasonal and spatial variability of sea surface temperature and chlorophyll-a within the Bay of Plenty. All the water quality, climatic and current data will be used to develop a productivity model which will allow assessments of the

sustainability of aquaculture within this region's coastal shelf waters. The modelling and assessments of sustainability are to be reported on by mid 2006.

Chapter 5: References

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Appendices

- Appendix I Site location and sampling program
- Appendix II BOP offshore transect survey results
- Appendix III CTD profiles for transect surveys

Appendix 1 – Site Location and Sampling Program

Sampling Sites

	Latitude Lor		NZMS 260 Grid ref. (m)		labstar #
Tauranga Tra	ansect				
10 m depth	37 35 40.9	176 08 34.8	2787737	6395999	730031
20 m	37 34 53.4	176 09 54.0	2789731	6397394	730032
30m	37 34 05.5	176 11 13.2	2791725	6398800	730033
50m	37 31 00.0	176 16 00.0	2798966	6404261	730034
100m	37 27 45.0	176 22 21.0	2808542	6409921	730035
200m	37 25 12.0	176 25 43.0	2813682	6414446	730036
Pukehina Tra	nsect				
10m	37 49 25.3	176 36 34.8	2827892	6369054	730043
20m	37 48 51.6	176 36 59.3	2828533	6370066	730044
30m	37 47 11.5	176 38 10.8	2830408	6373078	730045
50m	37 45 05.0	176 39 41.1	2832774	6376884	730046
100m	37 40 12.4	176 43 10.9	2838278	6385687	730047
200m	37 38 30.7	176 44 23.6	2840189	6388745	730048
Whakatane T	ransect				
10m	37 56 02.0	177 00 53.3	2862982	6355319	730037
20 m	37 55 35.4	177 01 17.3	2863604	6356112	730038
30m	37 53 43.1	177 02 22.4	2865348	6359500	730039
50m	37 51 41.9	177 04 00.0	2867899	6363125	730040
100m	37 44 12.9	177 08 30.5	2875141	6376649	730041
200m	37 41 51.9	177 10 02.0	2877580	6380891	730042
Opotiki Trans	ect				
10m	37 58 41.4	177 15 44.1	2884479	6349415	730049
20m	37 57 50.0	177 15 47.5	2884636	6350990	730050
30m	37 56 04.3	177 15 58.0	2885047	6354233	730051
50m	37 51 55.7	177 16 23.1	2886021	6361859	730052
100m	37 44 42.6	177 17 06.2	2887710	6375147	730053
200m	37 39 27.4	177 17 37.3	2888933	6384817	730054

Sampling program

At each sampling station the following parameters were recorded; vertical light attenuation, secchi depth, and dissolved oxygen, temperature, fluorescence, turbidity and salinity/conductivity profiles. These profiles are based on the dow n-cast recordings from the SBE 19 Plus probe.

Surface sampling (0-5m) in the plans below is an integrated tube sample except for the Enterococcal samples.

Sampling plan for phytoplankton identification and enumeration, 168 spot samples (14 per transect)

Sample Sampling Station (depth contour m)

Depth (m)	10	20	30	50	100	200
0-5.	х	х	х	х	•	Х
10						
20		х	х	х		Х
30				х		Х
50				х		х
100				х		
200						

Sampling plan for phytoplankton size fractionation (Chl-a), 144 spot samples (9 per transect) Sample Sampling Station (depth contour m)

				\		,
Depth(m)	10	20	30	50	100	200
0-5 .		х		х		х
10						
20		х		х		
30				х		х
50				х		
100				х		
200						

Sampling plan for dissolved nutrients DRP, NH4, NO3, 288 spot samples (18 per transect) Sample Sampling Station (depth contour m)

Depth(m)	10	20	30	50	100	200		
0-5.	х	х	х	х	х	Х		
10								
20	х	х	х		х			
30			х	х		Х		
50				х	х	х		
100					х			
200						Х		

Sampling plan for TN, TP, SS, DRSi, DFe and TOC/DOC, 192 spot samples (16 per transect).

Sample	Sam	pling S	station	(aepth d	contour	m)
Depth(m)	10	20	30	50	100	200
0-5.	Х	х	х	х	х	х
10				•	•	
20		х	х	х		
30			х	Х		х
50				х	х	
100					х	
200						х

Sampling plan for Enterococci bacteria, 96 spot samples (6 per transect).

Sample Sampling S	tation ((depth c	contour	m)		
Depth(m)	10	20	30	50	100	200
0	х	х	Х	х	х	х

Appendix 2 – BOP Offshore Transect Survey Results

In the follow ing results the first column lists the transect sampled (Pukehina, Whakatane & Opotiki) with the depth contour along the transect given in the fourth column, then the actual water depth at that site from which the samples were taken. The follow ing abbreviations and units are used;

Temp	temperature (degrees Celsius)
Cond	conductivity @ 25 °C
Saln	salinity
Secchi	secchi disc (measured in metres without a viewing tube)
Vlec	vertical light extinction coefficient derived from PAR measurements
Ent	Enterocci number cfu/100ml
PH	pH measurement @ 25°C
SS	Suspended Solids (g/m ³)
TOC	Total Organic Carbon (g/m ³)
DOC	Dissolved Organic Carbon (g/m ³)
DNPOC	Dissolved non-purgable organic carbon (g/m ³)
NPOC Non-pu	ırgable organic carbon (g/m³)
DRSi	Dissolved Reactive Silica (mg/m ³)
DFe	Dissolved iron (mg/m³)
TN	Total nitrogen (mg/m ³)
NH4	Ammonium nitrogen (mg/m ³)
NOx	Oxidised nitrogen (mg/m ³)
TP	Total Phosphorus (mg/m ³)
DRP	Dissolved Reactive Phosphorus (mg/m ³)
Chla	Chlorophyll-a total filterable on 0.7µm (mg/m ³)
Nanochla	Chlorophyll-a for plankton 0.7-20µm (mg/m³)
Microchla	Chlorophyll-a for plankton 20-200 µm (mg/m ³)
Plankton	Phytoplankton (thousands of cells per litre)

ankton	121.5	60.0	210.0		234.0	286.0 521 0	20120	2		8.0	326.0	485.0 543.0	120.5		104.7	159.0	32.U	139.3 481 0	2	203.0	408.5	171.0	24.5		15.0	309.0	249.5	280.0	15.3	13 E	151.0	21.7	158.6	601.0		310./ 212.0	53.0	3.0			8.0	352.0 675.0	443.0	188.5	
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lankton	207.5	218.5	34.3	165.0	273.0		90.3	0.1.0 20.0	50	5	1.8	3.8	1.0	с с	109.0	224.0	103.5	112.3	201.5		91.5 46.0	125.0	54.5	1	0.0 0	ν.α V	3.7	300.0	186.0	145.5	163.5		223.0	151.0	108.5	43.0		5.3	13.0	14.4
Aicroch ol	-	0.66	0.46				0.48	0.40	7C.0		0.46		0.4	200	500	0.59	0.61			1	0.0 7 0	0.5	0.31		0.4	0.4	0.24		1.12	1.18			0.86	0.99	0.64	0.62		0.36	0.34	0.58
VanoCh N		154	0.81				1.81	0.0	10		0.07		0.07	500	200	1.39	2.67				1.14 0.86	1.03	0.59		0.05	0.04	0.03		1.73	1.79			2.06	1.74	1.38	0.95		0.09	0.12	0.09
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ec Ent pH SS TO	22 8.1 11	23 8.1 16	8 22	18 8.1 20	8.1 23	8.1 20	15 8.1 22 8.1 22	0 20 8 1 - 21	8.1 22	08 8.1 20	8.2 18	8.1 24	8.1 9.1 20	0.1 8 27	16 0.5 8.1 9	22 0.5 8.1 20	8.1 22	19 0.5 8.1 16	8.1 17	42 DE 01 1/	1.2 U.3 0.1 3.4 8.1 15	8.1 15	8.1 17	09 0.5 8.1 15 6.1 15	8.1 8 0	8.1 16 8.1 16	8.1 20	30 2.0 8.2 9	26 1.0 8.2 5.8	40 0E 02 7.4	10 U.O 0.2 1.4 8.2 5.6	8.2 4.8	19 1.0 8.2 7	8.2 6.6	8.2 4.8	8.1 4	10 0.5 8.1 2.8	8.2 3.6	8.1 4.8	8.2 6.6 7.7 6
ichi Viec Ent pH SS TO	4.2 0.22 8.1 11	6.3 0.23 8.1 16	8 22	8.6 0.18 8.1 20	8.1 23	8.1 20	9.2 0.15 8.1 22 8.1 20	0.1 20 8.1 21	8.1 22	8.2 0.08 8.1 20	8.2 18	8.1 24	8.1 10 e 1 20	0 20 8 22	5.2 0.16 0.5 8.1 9	6.8 0.22 0.5 8.1 20	8.1 22	8.2 0.19 0.5 8.1 16	8.1 17	10 012 0F 01 01	1.0 0.13 0.3 0.1 3.4 8.1 15	8.1 15	8.1 17	0.1 0.09 0.5 8.1 15	0.1 0 0 0	8.1 16 8.1 16	8.1 20	6.3 0.30 2.0 8.2 9	6.2 0.26 1.0 8.2 5.8 2.2 0.26	8.7 8.7	0./ U.IO U.O 0.2 0.4	8.2 4.8	3.1 0.19 1.0 8.2 7	8.2 6.6	8.2 4.8	8.1 4	8.1 0.10 0.5 8.1 2.8	8.2 3.6	8.1 4.8	8.2 6.6 7.7 6
n Secchi Vlec Ent pH SS TO	5 4.2 0.22 8.1 11	8 6.3 0.23 8.1 16	6 8 22	.2 8.6 0.18 8.1 20	4 8.1 23	8.1 20	.5 9.2 0.15 8.1 22 3 8.1 22	8 81 21	9 8.1 22 9	8 18.2 0.08 8.1 20	.8 8.2 18	.8 .1 24	./ 8.1 10 6 8.1 20	0 0.1 20	5.2 0.16 0.5 8.1 9	6.8 0.22 0.5 8.1 20	8.1 22	8.2 0.19 0.5 8.1 16	8.1 17	8.1 1/ 110 012 05 01 01	11.0 0.13 0.3 0.1 9.4 8.1 15	8.1 15	8.1 17	9 10.1 0.09 0.5 8.1 15	0.1 0 0 0 0	9 8.1 16	8.1 20	2 6.3 0.30 2.0 8.2 9	4 6.2 0.26 1.0 8.2 5.8	9 9 9 9 9 9 9 7 9 7 9 7 9 7 9 7 9 7 9 7	0 0./ 0.10 0.0 0.2 7.4 1 8.2 5.6	2 8.2 4.8	2 13.1 0.19 1.0 8.2 7	4 8.2 6.6	5 8.2 4.8	7 8.1 4	9 8.1 0.10 0.5 8.1 2.8	9 8.2 3.6	9 8.1 4.8	8 8.2 6.6 4 7.7 6
d Saln Secchi Vlec Ent pH SS TC	30 31.5 4.2 0.22 8.1 11	0 33.8 6.3 0.23 8.1 16	50 34.6 8 22	0 34.2 8.6 0.18 8.1 20	0 34.4 8.1 23	0 34.8 8.1 20	0 34.5 9.2 0.15 8.1 22	0 348 81 21	0 34.9 8.1 22	0 34.8 18.2 0.08 8.1 20	0 34.8 8.2 18	0 34.8 8.1 24	0 34.7 8.1 10 0 34.6 8.1 20	0 35.0 8.1 20	5.2 0.16 0.5 8.1 9	6.8 0.22 0.5 8.1 20	8.1 22	8.2 0.19 0.5 8.1 16	8.1 17	8.1 1/ 8.1 0.12 0.5 0.1 0.1	11.0 0.13 0.3 0.1 9.4 8.1 15	8.1 15	8.1 17	0 34.9 10.1 0.09 0.5 8.1 15	0 34.9 8.1 18 0 34.0 8.1 0	0 34.9 8.1 16	0 34.8 8.1 20	7 33.2 6.3 0.30 2.0 8.2 9	0 33.4 6.2 0.26 1.0 8.2 5.8	0 33.9 8.7 8.2 8.2 8.2 0 32.6 8.7 8.2 0 32.6 8.7 8.2 8.2 8.2 8.2 8.2 8.2 8.2 8.2 8.2 8.2	0 34.1 0.7 0.10 0.3 0.2 7.4 0 34.1 8.2 5.6	0 34.2 8.2 4.8	0 34.2 13.1 0.19 1.0 8.2 7	0 34.4 8.2 6.6	0 34.5 8.2 4.8	0 34.7 8.1 4	0 34.9 8.1 0.10 0.5 8.1 2.8	0 34.9 8.2 3.6	0 34.9 8.1 4.8	0 34.8 8.2 6.6 0 34.4 7.7 6
p Cond Saln Secchi Vlec Ent pH SS TC	4 5080 31.5 4.2 0.22 8.1 11	5 5140 33.8 6.3 0.23 8.1 16	4 5250 34.6 8 22	7 5190 34.2 8.6 0.18 8.1 20	0 5210 34.4 8.1 23	3 5270 34.8 8.1 20	0 5230 34.5 9.2 0.15 8.1 22 1 5210 343 8.2 0.15 8.1 22	4 5270 34.8 8.1 21	4 5280 34.9 8.1 22	2 5270 34.8 18.2 0.08 8.1 20	1 5270 34.8 8.2 18	1 5270 34.8 8.1 24	1 2260 34./ 8.1 10 6 5240 34.6 8.1 20	5 5300 35.0 x 20	2 2 2 2 0 10 0 10 0 2 8 1 9	8 6.8 0.22 0.5 8.1 20	5 8.1 22	9 8.2 0.19 0.5 8.1 16	0 8.1 17		2 8.1 15	4 8.1 15	8.1 17	0 5290 34.9 10.1 0.09 0.5 8.1 15 0 5200 34.0 10.1 0.09 0.5 8.1 15	0 0290 04.9 8.1 18 0 5000 34.0 8.1 0	0 5280 34.9 8.1 16 8.1 16	8 5280 34.8 8.1 20	9 4957 33.2 6.3 0.30 2.0 8.2 9	5 5140 33.4 6.2 0.26 1.0 8.2 5.8	0 01/U 33.9 87 018 05 87 74	5 5190 34.1 0.7 0.10 0.3 0.2 7.4	7 5200 34.2 8.2 4.8	3 5210 34.2 13.1 0.19 1.0 8.2 7	3 5220 34.4 8.2 6.6	1 5230 34.5 8.2 4.8	1 5260 34.7 8.1 4	5 5290 34.9 8.1 0.10 0.5 8.1 2.8	5 5290 34.9 8.2 3.6	5 5290 34.9 8.1 4.8	5210 34.4 8.2 6.6 t 5210 34.4 7.7 6
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Appendix 3 – CTD Profiles From Transect Surveys

CTD transect data covers the period from October 2003 up to December 2004.

Graph titles start with the transect;

"Opo" = Opotiki, "Whk" = Whakatane, "Puk" = Pukehina.

Next in graph title is the depth contour and then the date of sampling. The bin.cnv can be ignored, although the "bin" indicates that that the graph is based on averaging the data over 1 m depth intervals.

For example "Opo20_171003bin.cnv" is data from the 20 m depth contour on the Opotikitransect sampled on the 17th October 2003.

In graphs Whk30_041203 and puk200_051203 there are what should be obvious fluorescence signal errors in the upper water column. On the August 2004 run of the transects the fluorometer was not functioning correctly so no data was obtained.

There are additional transects profiled for $7-9^{th}$ December 2004 labelled T05 – T15, and these correspond to the ASR Ltd shelf sediment survey as follow s;

- T05 = "C" and is off Matata
- TO6 = "D" and is off Thornton
- T11 = "I" and is off Opape
- T13 = "K" and is off Omaio Bay
- T15 = "M" and is off Raukokore







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58

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